

Role of Obestatin in Improvement of Obesity-Induced Metabolic and Kidney Function Changes in Exercised Rats

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Abstract

Background: Obesity is a common health problem which may be caused by feeding a high fat diet (HFD) and has caused many complications including metabolic and kidney function changes. On the other hand, obestatin is an adipokine that was found to be expressed also in the kidney and decrease body weight of obese rats. Also, moderate exercise training was suggested to improve obesity and minimize its complications, but, the exact mechanism was not well understood.

Aim: To investigate the obesity-induced metabolic and kidney function changes in exercised rats and clarify the potential role of obestatin.

Material and Methods: 48 male albino rats of local strains were randomly divided into two groups (I and II). Group I (lean group, n=24) included group IA (sedentary group, n=8), group IB (exercised group, n=8), and, group IC (obestatin treated group, n=8). Group II [obese, high fat diet (HFD)-induced obesity, n=24] was formed of group IIA (obese Sedentary, n=8), group IIB (obese exercised, n=8), and, group IIC (obestatin treated, n=8). At the end of experimental period, blood samples were obtained and allowed to clot at room temperature before centrifugation to obtain serum that was stored at -20 ° C for biochemical assessment.

Results: Obese groups (sedentary, exercised and obestatin treated) had a significant increase in body mass index (BMI), adiposity %, serum [glucose, insulin, C-peptide, total cholesterol (TC), triglycerides (TG), low-density lipoprotein (LDL), C-reactive protein (CRP), tumor necrosis factor alpha (TNF α), creatinine, urea and K⁺], urinary [Na⁺, total protein and albumin], homeostasis model assessment of insulin resistance index (HOMA-IR) and urine volume, with a significant decrease in serum [obestatin, high-density lipoprotein (HDL), Na⁺, total protein and albumin] and urinary (creatinine and K⁺) and creatinine clearance in the same groups. On the other hand, obese exercised and obese obestatin treated groups showed a significant reduction in BMI, adiposity %, serum (glucose, insulin and C-peptide, TC, TG, LDL, CRP, TNF α creatinine, urea and K⁺), urinary (Na⁺, total protein, albumin) and urine volume, with a significant

increase in serum (obestatin, HDL, Na⁺, total protein and albumin) and urinary (creatinine and K⁺) and creatinine clearance in the same groups. Within the obese sedentary and obese exercised groups, serum obestatin was negatively associated with BMI, atherogenic index, serum (glucose, insulin, TC, TNF α , creatinine and urea), HOMA-IR, and urinary Na⁺, but, it was positively associated with Homeostatic Model Assessment of beta cell function (HOMA-B), urinary K⁺ and creatinine clearance.

Conclusion: Moderate exercise training improved both metabolic and kidney function changes that occurred with HFD-induced obesity through its anti-inflammatory effect and increasing obestatin secretion.

Keywords: Obesity, Exercise training, Obestatin, Kidney function.

1. Introduction

Obesity is a chronic disease that was defined as increased body fat accumulation, and, it was associated with metabolic abnormality ^[1]. Park, Kim ^[2] and Soltani, Washco ^[3] declared that excessive caloric intake contributed to obesity and initiated a cascade that altered kidney functions and finally lead to obesity-related chronic kidney disease.

Previous studies declared that adipose tissue produced bioactive substances termed adipokines ^[4] which induced production of reactive oxygen species (ROS) causing systemic oxidative stress that increased production of proinflammatory cytokines causing renal injury ^[5]. On the other hand, Teixeira-Lemos, Nunes ^[6] proved that exercise training reduced oxidative stress.

23-amino acid-peptide, Obestatin is. а synthesized from preproghrelin which also gives rise to ghrelin ^[7]. It is produced by gastric mucosa, duodenum, pancreas, kidney, brain, testis. mammary glands and adipose tissue ^[4, 7-9]. Grönberg, Tsolakis ^[10] confirmed that obestatin was expressed by both pancreatic islets and ducts. Previous studies showed controversies about effect of obestatin on body metabolism. Green and Grieve ^[11] stated that obestatin had positive effects on pancreatic β -cells and islets, while, Agnew, Calderwood ^[12] and Unniappan, Speck ^[13] declared that administration of obestatin to normoglycemic rats had no effect on circulating glucose or insulin.

On the other hand, Ren, Guo ^[14] found that obestatin inhibited glucose-induced insulin secretion. Also, Gao, Kuang ^[15], Lippl, Erdmann ^[16], Egido, Hernández ^[17] and Zhang, Li ^[18] reported an inverse relationship between circulating obestatin and insulin levels. On the contrary, Granata, Settanni ^[19] and Pradhan, Wu ^[20] confirmed the ability of obestatin to stimulate insulin secretion under hyperglycemic conditions. On the other hand, Agnew, Calderwood ^[12] and Nagaraj, Raghavan ^[21] stated that chronic obestatin treatment reduced blood triglycerides with no effect on blood cholesterol. On the other hand, Gao, Kuang ^[15], Gao, Kuang ^[22], Shen, Yu ^[23] and Wang, Li ^[24] detected a decrease in blood obestatin in obese patients with hyperglycemia, metabolic syndrome, and insulin resistance.

Also, Lee, Chen ^[25] confirmed that obese patients who achieved body weight reduction had increased obestatin levels. On the contrary, Arrigo, Gitto ^[26] and Prodam, Cadario ^[27] found increased obestatin levels in patients with obesity, whilst Lee, Chen ^[25] and Siejka, Jankiewicz-Wika ^[28] stated that weight loss in obese patients had no effect on obestatin levels. Also, up to our knowledge, there was no study that assessed effect of moderate exercise training and exogenous obestatin treatment on kidney function changes in case of diet-induced obesity.

So, this study was conducted to investigate the obesity-induced metabolic and kidney function changes occurred in exercised rats and to clarify the potential role of obestatin.

2. Materials and Methods

In Zagazig Faculty of Medicine Physiology Department, this study was achieved, from July 2018 to December 2018. Forty-eight adult healthy male albino rats, of local strains, weighing 180 -220 gm were obtained from the animal house of Faculty of Medicine, Zagazig University. They were put in steel wire cages (four per cage) in the Physiology Department research laboratory, under hygienic conditions, at room temperature $(23 \pm 3^{\circ} C)$ and on 12-hour light/dark cycle. For accommodation to laboratory conditions, rats were kept for one week before starting the experimental program on free access to commercial rat laboratory chow and water.

The used experimental protocol in this study was achieved according to the data guiding the use of research animals and was approved by Institutional Research Board of Faculty of Medicine, Zagazig University.

Experimental design: The rats were randomly divided into two groups I (lean) and II (obese). Group I (lean group, n=24): the animals in this group were fed on ordinary laboratory chow diet (consisted of carbohydrate 62.8%, protein 25.8% and fat 11.4% and it was obtained from Zagazig Faculty of Agriculture) for 14 weeks (duration of the study), and further subdivided into 3 subgroups; group IA (sedentary group, n=8) in which the rats remained sedentary in their cages (no exercise), group IB (exercised group, n=8) in which the rats were trained by swimming exercise training of moderate intensity (one hour/day, five days/week) ^[29] in the last 6 weeks of this 14-week-study, and, group IC (obestatin treated group, n=8) in which

rats received a single daily i.p. dose of obestatin (64 µg/kg body weight dissolved in 0.1 ml normal saline) ^[30] for the last 6 weeks of the study while they remained sedentary in their cages. Rats of lean sedentary and lean exercised groups were injected i.p. daily with a single dose of 0.1 ml normal saline in the last 6 weeks of the study. Obestatin was purchased from Sigma Aldrich Co.-USA (Catalog No. 00266). Group II [obese, high fat diet (HFD)induced obesity, n=24]: rats in this group were fed a HFD for 8 weeks; this HFD consisted of (16.4% protein, 25.6% carbohydrate and 58.0% fat in the form of cotton seed oil added to the laboratory chow diet [31] and further subdivided into 3 subgroups; group IIA (obese Sedentary, n=8) in which the rats remained sedentary in their cages (no exercise), group IIB (obese exercised, n=8) in which the rats were trained by swimming exercise training of moderate intensity in the last 6 weeks of the study as in group IB, and, group IIC (obese obestatin treated, n=8) in which the rats received a single daily i.p. dose of obestatin, as in group IC, for the last 6 weeks of the study while they remained sedentary in their cages. Rats of obese sedentary and obese exercised groups were injected i.p. daily with a single dose of 0.1 ml normal saline in the last 6 weeks of the study.

	Lean sedentary (8 rats)	Ordinary diet	Ordinary diet + Saline		
Lean group (24 rats)	Lean exercised (8 rats)	Ordinary diet	Ordinary diet + Exercise +		
			Saline		
	Lean obestatin treated (8 rats)	Ordinary diet	Ordinary diet + Obestatin		
Obese group (24 rats)	Obese sedentary (8 rats)	HFD	HFD + Saline		
	Obese exercised (8 rats)	HFD	HFD + Exercise + Saline		
	Obese obestatin treated (8 rats)	HFD	HFD + Obestatin		
Duration (14 weeks)		8 weeks	6 weeks		

Table 1: Experimental design

HFD= High fat diet.

Body mass index (BMI) was calculated at the end of the 8th week (as an indicator of obesity if it was more than 0.68 gm/ cm² ^[32]), and at the end of the study (to explore the changes in BMI among different groups).

This equation was used:

BMI= [Body weight (gm)] / [Length² (cm²)(from nose to anus length)]^[32]

Swimming exercise program: The rats in the exercised groups performed moderate intensity

swimming exercise, one hour/day, five days/week for 6 weeks. Swimming was practiced in a cylindrical tank of 45 cm diameter, 80 cm high and filled with 32- 35 ° C water 45 cm deep ^[33]. Swimming rats were initially trained for 15 minutes/day and duration was gradually increased such that the rats were able to perform exercise for one hour/day, which was achieved in one week ^[34]. Exercise was performed between 9- 10 am. At the end of each exercise session, the animals were kept to dry in a warm environment ^[35]. The animals that practiced exercise were sacrificed 48 h after the end of the last training session to minimize the acute effects of the exercise ^[6].

At the end of experimental period, all rats were placed individually in metabolic cages for 24 hr. Urine was collected, centrifuged and stored at -20° C to avoid urea degradation ^[36] until analysis.

After 12 hours fasting, the blood samples were obtained from all rats after their scarification by decapitation. Blood samples were allowed to clot at room temperature $(23 \pm 3^{\circ} \text{ C})$ before centrifuging for 15 minutes at 3000 revolutions per minute (rpm). The separated serum was stored at -20° C in dark containers for biochemical assay.

Serum levels of obestatin, insulin, glucose, triglycerides (TG), total cholesterol (TC), highdensity lipoprotein (HDL), tumor necrosis factor alpha (TNF α), C-reactive protein (CRP), creatinine and urea were measured using commercial kits from Sigma-Aldrich, USA (Cat. No. RAB0208. RAB0904, GAGO20, TR0100, MAK043, MAK045, RAB0479, RAB0097, MAK080 and MAK006, respectively). Also, commercial kits were used for estimating serum levels of C-Peptide (ALPCO, Cat. No. 80-CPTRT-E01), total protein (Biocompare.com, Cat. No. ABIN996403) and albumin (abcam.com, Cat. No. ab235642). Moreover, commercial kits were used for estimating serum and urine levels of Na⁺ (abcam.com, Cat. No. ab211096) and K⁺ (Sigma-Aldrich, USA, Cat. No. 37202). Furthermore, commercial kits were used for measuring the urinary levels of creatinine (Sigma-Aldrich, USA, Cat. No. MAK080), total protein (Chondrex.com, Cat. No. 9040) and albumin (abcam.com, Cat. No. ab235642).

Homeostasis model assessment of insulin resistance (HOMA-IR) index was measured by the formula:

HOMA-IR= [(Insulin in μ IU/L) X (Glucose in mg/dl)] / 405^[37]

Homeostatic Model Assessment of beta cell function (HOMA-B) index was measured by the formula:

HOMA-B = $[(20 \text{ X Fasting Insulin in } \mu \text{IU/L}) / (\text{Fasting Glucose in mmol/L} - 3.5)]^{[38]}$

Low-density lipoprotein (LDL) serum level was calculated using Friedewald formula:

LDL $(mg/dl) = [TC] - [(HDL) + (TG / 5)]^{[39]}$

Atherogenic index was calculated using the formula:

Atherogenic index= $[TC - HDL] / HDL^{[40]}$

Adiposity index (%) was determined by the sum of epididymal, visceral and retroperitoneal fat weights divided by body weight $x100^{[41]}$.

Calculation of creatinine clearance:

Creatinine clearance (ml/min) = [(Urine Creatinine in mg/dl) X (Urine Volume in ml/day)] / [(Serum Creatinine in mg/dl) X (Time in min)]^[42]

Statistical Analysis: The obtained data was expressed as mean values \pm standard deviation (Mean \pm SD). Means were compared by one-way analysis of variance (ANOVA) and Tukey HSD for Post hoc Multiple Comparisons using (IBM SPSS Statistics Version 25 Software for Windows) for statistical significance. Pearson correlation analysis using (GraphPad Prism Version 7 Software for Windows) was performed to study the associations between serum obestatin and different studied parameters within obese sedentary and obese exercised groups. P value \leq 0.05 indicated significance.

3. Results

In this study, we investigated the effects of both exercise training and obestatin treatment on both metabolic and kidney function changes in dietinduced obesity. Obesity was induced by giving rats HFD for 8 weeks in the obese groups, then, rats of obese exercised group were exposed to a moderate exercise and those of obese obestatin treated group were administered obestatin for extra 6 weeks.

In Table 2, there was a significant (P<0.001) increase in both BMI and adiposity % in obese groups (sedentary, exercised and obestatin treated), in comparison to those of the lean groups. On the other hand, with either, performing exercise training (in obese exercised group) or treatment with obestatin (in obese obestatin treated group), for next 6 weeks, both BMI and adiposity % were significantly (P<0.05) reduced in comparison to obese sedentary group. Changes in serum obestatin showed a significant (P<0.01) increase in lean obestatin treated group in comparison to lean sedentary group.

On the other hand, a significant (P < 0.001) reduction was detected in serum obestatin in obese groups (sedentary, exercised and obestatin treated), in comparison to those of the lean groups. On the contrary, a significant (P<0.01) increase in serum obestatin occurred in both obese exercised and obese obestatin treated groups in comparison to the obese sedentary group. Also, a significant (P<0.001) increase was found in serum level of glucose, insulin and C-peptide, and, HOMA-IR in obese groups (sedentary, exercised and obestatin treated), in comparison to those of the lean groups. On the other hand, a significant (P<0.001) decrease in serum level of both glucose and C-peptide occurred in both obese exercised and obese obestatin treated groups in comparison to those in the obese sedentary group.

Also, a significant (P<0.01) decrease in serum insulin was detected in both obese exercised and obese obestatin treated groups in comparison to that in the obese sedentary group. In comparison to obese exercised group, a significant (P<0.05) increase in serum level of insulin, C-peptide and HOMA-IR was found in obese obestatin treated group. On the other hand, a significant (P<0.05) decrease was found in HOMA-B in obese groups (sedentary, exercised and obestatin treated), in comparison to that of the lean groups.

In comparison to lean groups (sedentary, exercised and obestatin treated), a significant (P<0.001) increase was found in serum level of TC, TG and LDL, but a significant (P<0.001) decrease was detected in serum HDL in obese groups. On the other hand, a significant (P<0.001) decrease was declared in serum level of TC, TG and LDL, but, a significant (P<0.01) increase was detected in serum HDL, in both obese exercised and obese obestatin treated groups in comparison to obese sedentary group.

Moreover, a significant (P<0.01) increase was found in atherogenic index in obese groups (sedentary, exercised and obestatin treated), in comparison to that in the lean groups. On the other hand, a significant (P<0.001) decrease was declared in atherogenic index in both obese exercised and obese obestatin treated groups in comparison to obese sedentary group. In comparison to lean groups (sedentary, exercised and obestatin treated), a significant (P<0.001) increase was found in serum CRP and TNF α in the obese groups. On the other hand, a significant (P<0.001) decrease was detected in serum CRP and TNF α in both obese exercised and obese obestatin treated groups in comparison to the obese sedentary group. But, a significant (P<0.05) increase was confirmed in serum CRP in obese obestatin treated group in comparison to obese exercised group.

In Table 3, kidney function changes among different groups were assessed. In comparison to lean groups (sedentary, exercised and obestatin treated), a significant (P<0.001) increase was detected in serum levels of creatinine, urea and K⁺, and in urinary Na⁺, total protein, albumin and urine volume, but, a significant (P<0.001) decrease was confirmed in serum levels of Na⁺, total protein and albumin, and, in urinary creatinine, K^+ , and, creatinine clearance, in obese groups. In comparison to obese sedentary group, a significant (P<0.001) decrease was found in serum levels of creatinine, urea, K⁺, and, in urinary Na⁺, total protein, albumin, and, urine volume, but, a significant (P < 0.05)increase was detected in serum levels of Na⁺, total protein, albumin, and, in urinary creatinine, K⁺, and, creatinine clearance, in both obese exercised and obese obestatin treated groups.

In comparison to obese exercised group, a significant (P<0.001) increase was estimated in serum creatinine, urea, K^+ , and, in urinary Na⁺, total protein, albumin, and urine volume, but, a significant (P<0.001) decrease was found in serum Na⁺, total protein, albumin, and, in urinary creatinine and K⁺, in obese obestatin treated group.

In Table 4, correlations within the obese sedentary group between serum obestatin and different studied parameters showed negative associations with final BMI (r=-0.959, P<0.001), serum glucose (r=-0.981, P<0.001), serum insulin (r=-0.964)P<0.001), HOMA-IR (r=-0.978)P<0.001), serum TC (r=-0.836, P<0.01), atherogenic index (r=-0.926, P<0.01), serum TNF α (r=-0.769, P<0.05), serum creatinine (r=-0.935, P<0.001), serum urea (r=-0.809, P<0.05) and urinary Na⁺ (r=-0.804, P<0.05), but, positive associations with HOMA-B (r=0.986, P<0.001), urinary K⁺ (r=0.729, P<0.05) and creatinine clearance (r=0.783, P<0.05).

Group	(Group I (Lean	.)	Group II (Obese)		
	Sedentary	Exercised	Obestatin	Sedentary	Exercised	Obestatin
Parameter	(IA)	(IB)	treated (IC)	(IIA)	(IIB)	treated (IIC)
BMI (gm/cm ²)	0.51±0.04	0.45±0.05	0.51±0.04	0.94±0.07 ^{a,b&c}	0.84±0.07 ^{a,b,c&}	0.81±0.06 ^{a,b,c&e}
					d	
Serum obestatin	157.9±6.7	161±9.1	172.6±8.6 ^f	105.9±6.6 ^{a,b&c}	123.4±6.4 ^{a,b,c&e}	132.9±9.7 ^{a,b,c&e}
(ng/ml)						
Serum glucose	86.6±9.3	73.9±6.8	81.4±7.6	296.3±19.4 ^{a,b}	228.4±17.3 ^{a,b,c}	240.3±18.9 ^{a,b,c}
(mg/dl)				&c	&g	&g
Serum insulin	12.2±1.8	9.5±1.2	11.1±1.8	35.8±2.9 ^{a,b&c}	25.9±2.4 ^{a,b,c&e}	29.7±3.6 ^{a,b,c,e&h}
(µIU/L)						
HOMA-IR	2.6 ± 0.5	1.7±0.3	2.2±0.4	26.1±2.5 ^{a,b&c}	14.6±1.7 ^{a,b,c&e}	17.6±2.4 ^{a,b,c,e&i}
HOMA-B	225.1±22.1	291.1±34.	261.1±29.8	55.7±7.3 ^{j,b&l}	57±7.7 ^{j,b&l}	61±10.1 ^{j,k&l}
		2				
Serum C-Peptide	104.1±11.3	86 ± 8.2^{f}	94.4±10.7	193±12.7 ^{a,b&c}	148.5±7.7 ^{a,b,c&g}	168.1±6.7 ^{a,b,c,g}
(ng/L)						&i
Serum TC (mg/dl)	180.9±10.9	169.3±10.	174±10.2	315±31.4 ^{a,b&c}	243.8±18.3 ^{a,b,c}	266.4±10.7 ^{a,b,c}
		7			&g	&g
Serum TG (mg/dl)	130.5±5.7	115.8±4.6	122.5±4.5	249.4±30.5 ^{a,b}	193.1±13.3 ^{a,b,c}	202±13.6 ^{a,b,c&g}
				&c	&g	
Serum HDL	47.12±2.7	60.6±3.1ª	53.6±4.2 ^{j&k}	25.1±3.6 ^{a,b&c}	38.5±4.3 ^{a,b,c&g}	33.5±4.1 ^{a,b,c&e}
(mg/dl)						
Serum LDL	107.7±10.7	85.5±11.8	95.9±11.2	240±30.1 ^{a,b&c}	166.6±19.9 ^{a,b,c}	192.5±12.4 ^{a,b,c}
(mg/dl)					&g	&g
Atherogenic index	2.8±0.3	1.8 ± 0.2	2.3±0.4	11.9±2.9 ^{a,b&c}	$5.4 \pm 1.1^{f,b,l\&g}$	7.1±1.2 ^{a,b,c&g}
Serum CRP	0.29±0.08	0.23±0.08	0.25±0.04	5.2±0.56 ^{a,b&c}	2.59±0.29 ^{a,b,c&g}	3.04±0.27 ^{a,b,c,g}
(µg/ml)						&h
Serum	21.1±1.5	17.3±1.7	18.8±1.4	82.2±7.7 ^{a,b&c}	48.4±5.4 ^{a,b,c&g}	52.5±5.1 ^{a,b,c&g}
TNF α (pg/ml)						
Adiposity %	5.4±0.4	4.8 ± 0.3^{f}	5.1±0.2	10.1±0.3 ^{a,b&c}	7.6±0.4 ^{a,b,c&g}	8.3±0.4 ^{a,b,c,g&i}

 Table 2: Biochemical changes among different groups

Data was expressed as Mean±SD. ^aP<0.001 in comparison to lean sedentary group. ^bP<0.001 in comparison to lean exercised group. ^cP<0.001 in comparison to lean obestatin treated group. ^dP<0.05 in comparison to obese sedentary group. ^eP<0.01 in comparison to obese sedentary group. ^fP<0.01 in comparison to lean sedentary group. ^gP<0.001 in comparison to obese sedentary group. ^hP<0.05 in comparison to obese sedentary group. ^hP<0.05 in comparison to obese exercised group. ⁱP<0.01 in comparison to obese sedentary group. ^hP<0.05 in comparison to obese exercised group. ⁱP<0.01 in comparison to lean obestatin treated group. BMI, body mass index; HOMA-IR, homeostasis model assessment of insulin resistance; HOMA-B, homeostasis model assessment of beta cell function; TC, total cholesterol; TG, triglycerides; HDL, high density lipoproteins; LDL, low density lipoproteins; CRP, C-reactive protein; TNF α , tumor necrosis factor alpha.

Group	Group I (Lean)			Group II (Obese)		
Group	Sedentar	Exercised	Obestatin	Sedentary	Exercised	Obestatin
Parameter	y (IA)	(IB)	treated	(IIA)	(IIB)	treated (IIC)
			(IC)			
Serum creatinine	0.62±0.0	0.56±0.0	0.59±0.0	2.11±0.06 ^{a,b&c}	1.53±0.11 ^{a,b,c}	1.86±0.09 ^{a,b,c,d&e}
(mg/dl)	3	2	3		&d	
Serum urea (mg/dl)	45±0.2	44.3±0.3	44.6±0.3	56.1±0.5 ^{a,b&c}	48.7±0.8 ^{a,b,c&d}	51±0.9 ^{a,b,c,d&e}
Serum Na ⁺ (mg/dl)	141.9±0.	141.5±0.	141.7±0.	122.7±0.7 ^{a,b&c}	134.7±0.6 ^{a,b,c}	129.5±0.9 ^{a,b,c,d&e}
	4	5	3		&d	

 Table 3: Kidney function changes among different groups

Serum K ⁺ (mg/dl)	2.73±0.0	2.66±0.0	2.72±0.0	5.58±0.15 ^{a,b&c}	3.36±0.16 ^{a,b,c}	4.2±0.2 ^{a,b,c,d&e}
	8	5	5		&d	
Serum total protein	6.37±0.1	6.35±0.1	6.34±0.1	4.57±0.23 ^{a,b&c}	5.61±0.17 ^{a,b,c}	5.09±0.16 ^{a,b,c,d&e}
(gm/dl)	4	7	8		&d	
Serum albumin (gm/dl)	2.91±0.1	2.91±0.1	2.9±0.09	1.55±0.08 ^{a,b&c}	2.44±0.05 ^{a,b,c}	2.01±0.09 ^{a,b,c,d&e}
	2	2			&d	
Urinary creatinine	41.4±0.5	41.1±0.3	40.9±0.2	28.5±0.7 ^{a,b&c}	38.4±0.7 ^{a,b,c&d}	35.6±0.8 ^{a,b,c,d&e}
(mg/dl)						
Urinary Na ⁺ (mg/dl)	132.6±1	133.1±0.	132.8±0.	148.5±0.4 ^{a,b&c}	137±0.6 ^{a,b,c&d}	139.7±0.5 ^{a,b,c,d&e}
		4	8			
Urinary K ⁺ (mg/dl)	126.4±0.	126.4±0.	126.1±0.	74.4±4.2 ^{a,b&c}	102.6±1 ^{a,b,c&d}	88.7±0.5 ^{a,b,c,d&e}
	6	6	6			
Urinary total protein	0.44±0.0	0.4±0.02	0.42±0.0	3.09±0.07 ^{a,b&c}	2.17±0.07 ^{a,b,c}	2.75±0.06 ^{a,b,c,d&e}
(gm/dl)	1		2		&d	
Urinary albumin	0.36±0.0	0.34±0.0	0.35±0.0	1.73±0.05 ^{a,b&c}	0.94±0.05 ^{a,b,c}	1.24±0.06 ^{a,b,c,d&e}
(gm/dl)	3	2	2		&d	
Urine volume (ml/24hr)	7.62±0.2	7.63±0.2	7.55±0.2	13.64±0.62 ^{a,b}	9.98±0.75 ^{a,b,c}	11.61±0.96 ^{a,b,c,d}
	1	1	1	&c	&d	&e
Creatinine clearance	0.36±0.0	0.39±0.0	0.36±0.0	0.13±0.01 ^{a,b&c}	0.17±0.01 ^{a,b,c}	0.16±0.02 ^{a,b,c&g}
(ml/min)	2	2f	2		&d	

Data was expressed as Mean±SD. ^aP<0.001 in comparison to lean sedentary group. ^bP<0.001 in comparison to lean exercised group. ^cP<0.001 in comparison to lean obestatin treated group. ^dP<0.001 in comparison to obese sedentary group. ^eP<0.001 in comparison to obese exercised group. ^fP<0.01 in comparison to lean sedentary group. ^gP<0.05 in comparison to obese sedentary group.

Table 4: Pearson's correlation coefficient (r) between serum level of obestatin and some biochemical
parameters in both obese sedentary and obese exercised groups

	Serum obestatin						
	Obese sede	entary group	Obese exercised group				
	r	Р	r	Р			
Final BMI	-0.959	< 0.001	-0.944	< 0.001			
Serum glucose	-0.981	< 0.001	-0.92	< 0.01			
Serum insulin	-0.964	< 0.001	-0.765	< 0.05			
HOMA-IR	-0.978	< 0.001	-0.97	< 0.001			
HOMA-B	0.986	< 0.001	0.712	< 0.05			
Serum TC	-0.836	< 0.01	-0.736	< 0.05			
Atherogenic index	-0.926	< 0.01	-0.955	< 0.001			
Serum TNFa	-0.769	< 0.05	-0.816	< 0.05			
Serum creatinine	-0.935	< 0.001	-0.72	< 0.05			
Serum urea	-0.809	< 0.05	-0.747	< 0.05			
Urinary Na ⁺	-0.804	< 0.05	-0.813	< 0.05			
Urinary K ⁺	0.729	< 0.05	0.79	< 0.05			
Creatinine clearance	0.783	<0.05	0.897	< 0.01			

P<0.05 indicated statistical significance. BMI, body mass index; HOMA-IR, homeostasis model assessment of insulin resistance; HOMA-B, homeostasis model assessment of beta cell function; TC, total cholesterol; TNF α tumor necrosis factor alpha.

On the other hand, correlations within the obese exercised group between serum obestatin and different studied parameters showed negative associations with final BMI (r=-0.944, P<0.001), serum glucose (r=-0.92, P<0.01), serum insulin (r=-0.765, P<0.05), HOMA-IR (r=-0.97, P<0.001), serum TC (r=-0.736, P<0.05), atherogenic index (r=-0.955, P<0.001), serum TNF α (r=-0.816, P<0.05), serum creatinine (r=-0.72, P<0.05), serum urea (r=-0.747, P<0.05) and urinary Na⁺ (r=-0.813, P<0.05), but, positive associations with HOMA-B (r=0.712, P<0.05), urinary K⁺ (r=0.79, P<0.05) and creatinine clearance (r=0.897, P<0.01).

4. Discussion

In this study, rats in the obese groups (sedentary, exercised and obestatin treated) were fed HFD to induce obesity that was confirmed by the use of BMI and adiposity % as a significant increase was found in their values in comparison to those of the lean groups. This was supported by EL-Gohary and Hussien ^[36] who declared that HFD-induced obesity was associated with increased BMI.

On performing exercise training (in obese exercised group) or treatment with obestatin (in obese obestatin treated group), for the last 6 weeks of the study, both BMI and adiposity % were significantly reduced in comparison to obese sedentary group. These results confirmed that training exercise and obestatin treatment deteriorated HFD-induced obesity which was supported by EL-Gohary and Hussien ^[36]. In contrary, Chow, Greenlund ^[43] declared that exercise alone is not effective for weight loss. This controversy may be referred to species difference, duration and type of exercise performed.

On the other hand, a significant reduction was detected in serum obestatin in obese groups (sedentary, exercised and obestatin treated), in comparison to that of the lean groups. Also, a negative correlation was estimated within the obese sedentary and obese exercised groups between serum obestatin and BMI. This was in agreement with Lacquaniti, Donato ^[44], Granata, Gallo ^[7] and Wang, Li ^[24] who confirmed that serum obestatin was inversely correlated with BMI. This may confirm that obesity had an inhibitory effect on obestatin production causing lack of its anorectic

effect leading to more weight gain. On the contrary, significant increases in serum obestatin levels occurred in both obese exercised and obese obestatin treated groups in comparison to the obese sedentary group.

These results declared that obestatin may have a role in exercise-induced modifications on metabolic and renal changes with obese rats. This was supported by Shen, Yu^[23] who declared that obestatin treatment decreased body weight by reduction of food intake, delay of gastric emptying, suppression of jejunal contraction and inhibition of ghrelin biological actions as it acted as an endogenous antagonist for ghrelin. Also, ^[18] stated that obestatin crossed blood-brain barrier and acted on brain centers to inhibit hunger. On the contrary, Wang, Chen^[45] stated that obestatin had no effect in exercise-induced change body weight. This controversy may be explained by the difference in type and duration of exercise training. Also, significant increases were found in serum levels of glucose, insulin and C-peptide, and in HOMA-IR in obese groups (sedentary, exercised and obestatin treated), in comparison to those of the lean groups.

Moreover, a negative correlation was found within the obese sedentary and obese exercised groups, between serum obestatin and serum glucose, serum insulin and HOMA-IR. These results increased insulin confirmed secretion and development of insulin resistance. This was supported by EL-Gohary and Hussien [36] and El Sawy, El-Sherbiny [46] who declared that HFDinduced obesity was associated with increased serum glucose, serum insulin and HOMA-IR. El Sawy, El-Sherbiny ^[46] explained the increased insulin level in HFD-induced obesity by fat accumulation in adipocytes, muscles and liver causing insulin resistance.

On the other hand, significant decreases in serum level of both glucose and C-peptide occurred in both obese exercised and obese obestatin treated groups in comparison to the obese sedentary group. This was in agreement with Saengsirisuwan, Perez ^[47] who confirmed the decrease in blood glucose with exercise and they owed this to increased glucose transport by increased GLUT-4 concentration in skeletal muscle and elevated activity of both hexokinase II and glycogen synthase enzymes. Also, Lacquaniti, Donato ^[44] confirmed that obestatin treatment improved glucose homeostasis. Also, a significant decrease in serum insulin was detected in both obese exercised and obese obestatin treated groups in comparison to the obese sedentary group. But, in comparison to obese exercised group, a significant increase in serum insulin was found in obese obestatin treated group.

Also, a significant increase was detected in both serum C-peptide and HOMA-IR in obese obestatin treated group in comparison to obese exercised group. These results confirmed that muscular exercise improved metabolic parameters more than obestatin treatment did. But, with exercise, obestatin serum levels were significantly increased which indicated that mechanism of actions of exercise was partly mediated through increasing serum obestatin levels. These results were supported by Savini, Catani^[48], EL-Gohary and Hussien [36], Gibb, Epstein [29] and EL-Goharv [49] confirmed that exercise training who deteriorated insulin resistance as evidenced by the decrease in plasma glucose, plasma insulin, and HOMA-IR. Gibb, Epstein ^[29] stated that exercise training increased both skeletal muscle glucose transporter (GLUT4) activity, and hepatic insulin clearance.

Also, Dewal, Stanford ^[50] declared that moderate exercise training increased glucose uptake by adipocytes. Moreover, Mafra, Guebre-Egziabher ^[51], Seim, Walpole ^[52] and Zhang, Li ^[18] declared that plasma obestatin was negatively associated with plasma insulin, plasma glucose and HOMA-IR. Furthermore, Granata, Gallo^[7] and Green and Grieve ^[11] confirmed that obestatin improved glucose tolerance and insulin sensitivity in HFDinduced obesity. On the other hand, a significant decrease was found in HOMA-B in obese groups (sedentary, exercised and obestatin treated), in comparison to those of the lean groups. Also, a positive correlation was detected within the obese sedentary and obese exercised groups between serum obestatin and HOMA-B. These results confirmed that obesity impaired beta cell function which was partially improved by exercise and obestatin treatment, although, this improvement was insignificant.

This was supported by Granata, Settanni ^[19], Favaro, Granata ^[53] and Green and Grieve ^[11] who

confirmed that obestatin improved β -cell function through enhancing its viability and inhibiting its apoptosis. In comparison to the lean groups (sedentary, exercised and obestatin treated), a significant increase was found in serum TC, TG and LDL, but a significant decrease was detected in serum HDL in obese groups. Also, a negative correlation was detected within the obese sedentary and obese exercised groups between serum obestatin serum TC. These results confirmed occurrence of impaired metabolic function with obesity. On the other hand, a significant decrease was declared in serum TC, TG and LDL in both obese exercised and obese obestatin treated groups in comparison to obese sedentary group.

On the contrary, a significant increase was detected in serum HDL in both obese exercised and obese obestatin treated groups in comparison to obese sedentary group. These results confirmed that exercise improved metabolic functions partially through increasing serum obestatin. These results were partially supported by El Sawy, El-Sherbiny ^[46] who found that obestatin treatment of obese rats decreased serum TC, serum TG and BMI, but they reported insignificant changes in both serum glucose and insulin levels. The partial difference in the results of that study may be related to the duration of treatment with obestatin. Mechanisms involved in improvement of lipid metabolism by obestatin include its ability to decrease food intake and body weight via reduction of perirenal and epididymal fat with reduction of stored fat ^[54]. Also, obestatin reduced intestinal absorption of TG^[12]. Moreover, obestatin increased fatty acid uptake, but, it inhibited lipolysis ^[44].

Furthermore, obestatin reduced serum lipids through stimulation of leptin secretion ^[46]. On the other hand, a significant increase was found in atherogenic index in obese groups (sedentary, exercised and obestatin treated), in comparison to those of the lean groups. This was in agreement with Ivanova, Myasoedova ^[55] who declared that HFD-induced obesity was manifested by increased blood LDL which passed through the endothelial fenestrations entering subendothelial spaces where it can be transformed into plaques that caused atherosclerosis and kidney disease progression. But, a significant decrease was reported in atherogenic index in both obese exercised and obese obestatin treated groups in comparison to obese sedentary group. Also, a negative association was found within the obese sedentary and obese exercised groups between serum obestatin and atherogenic index.

These changes in atherogenic index with obese exercised and obese obestatin treated groups can be explained by the improvement in the lipid profile that occurred in these groups in comparison to the obese sedentary group. In comparison to lean groups (sedentary, exercised and obestatin treated), a significant increase was found in serum levels of both CRP and TNF α in the obese groups. Also, a negative correlation was declared within the obese sedentary and obese exercised groups between serum levels of both obestatin and $TNF\alpha$. These results confirmed presence of inflammatory reaction with obesity. This was supported by Speretta, Rosante ^[56] who explained the obesity-induced insulin resistance by the increase in serum TNFa increased insulin resistance which through increasing the release of free fatty acids in adipocytes and by blocking the synthesis of adiponectin.

On the other hand, a significant decrease occurred in serum CRP and TNF α in both obese exercised and obese obestatin treated groups in comparison to obese sedentary group. These results confirmed that both exercise and obestatin had antiinflammatory activities. This was supported by Speretta, Rosante ^[56] and EL-Gohary and Hussien ^[36] who declared that $TNF\alpha$ was reduced among obese exercised rats. Also, Savini, Catani ^[48] stated that exercise had antioxidant and anti-inflammatory obesity-associated effects that prevented complications through improving glucose homeostasis and antioxidant body defenses.

In comparison to lean groups (sedentary, exercised and obestatin treated), a significant increase was detected in serum levels of creatinine, urea and K^+ , and in urinary Na⁺, total protein, albumin and urine volume in obese groups. Also, a negative correlation was detected within the obese sedentary and obese exercised groups between serum obestatin and each of serum creatinine, serum urea and urinary Na⁺. On the other hand, a significant decrease was detected in serum levels of Na⁺, total protein and albumin, and, urinary creatinine, K⁺, and, creatinine clearance in obese

groups (sedentary, exercised and obestatin treated) in comparison to lean groups.

Also, a positive correlation was found within the obese sedentary and obese exercised groups between serum obestatin and each of urinary K⁺ and creatinine clearance. These results confirmed that obesity deteriorated kidney function and this was accompanied by low obestatin level. This was supported by Lacquaniti, Donato^[44] who found low obestatin levels in chronic kidney disease. Also, Amin, Kamel^[57] and EL-Gohary and Hussien^[36] declared that obese rats had a significant elevation in serum urea, serum creatinine, and urinary albumin. Moreover, Soltani, Washco ^[3] explained the disturbances occurred in electrolytes and nitrogenous waste products with kidney disease by the essential role exerted by the kidney in regulation of these substances.

Furthermore, Wang, Chen ^[58] supported the deterioration in kidney function that occurred with obesity and they owed this to the binding of the elevated glucose irreversibly to proteins in the kidney forming advanced glycosylation end products that finally stimulated fibrotic growth factors causing renal damage. In addition, Kume, Uzu ^[59] stated that HFD caused an imbalance between lipolysis and lipogenesis in the kidney in addition to the systemic metabolic changes that finally lead to renal lipid accumulation with subsequent kidney injury. This was supported by Choudhary, Naheed ^[60] who confirmed that hyperlipidemia was associated with renal function deterioration.

Also, Tokuyama, Wakino [61] declared that obesity increased glomerular injury and caused glomerular hyperfiltration. Moreover, Serra, Romero ^[62] confirmed that prolonged obesity was manifested by proteinuria. Furthermore, El-Wakf, Serag ^[63] stated that HFD-induced obesity caused kidney dysfunction that was characterized by changes in both serum and urine of Na⁺ and K⁺. Soltani, Washco ^[3] explained the presence of proteinuria and changed serum and urinary Na⁺ and K^+ with obesity-induced kidney disease by occurrence of glomerular membrane degeneration, and renal tubular injury.

On the other hand, Darouich, Goucha ^[64] detected occurrence of renal injury with obesity and they owed this to oxidative stress that has been

detected in obesity-related renal diseases. EL-Gohary and Hussien ^[36] explained occurrence of obesity-induced oxidative stress by the increased adipose tissue which enhanced proinflammatory cytokines as TNF α that increased production of reactive oxygen species. This was agreed with Fernández-Sánchez, Madrigal-Santillán ^[5] who stated that obesity was considered as a state of chronic inflammation as adipocytes produced proinflammatory cytokines including TNF α .

Also, Noeman, Hamooda^[41] declared that renal lipids are rich in long-chain polyunsaturated fatty acids which made kidney to be more liable to damage by ROS which oxidized lipids and proteins causing cellular injury and increasing glomerular and renal tubule damage and proteinuria ^[65]. In comparison to obese sedentary group, a significant decrease was found in serum levels of creatinine, urea, K⁺, and, in urinary Na⁺, total protein, albumin, and, urine volume in both obese exercised and obese obestatin treated groups. But, a significant increase was found in serum levels of Na⁺, total protein, albumin, and, in urinary creatinine, K⁺, and, creatinine clearance in both obese exercised and obese obestatin treated groups, in comparison to obese sedentary group.

Also, in comparison to obese exercised group, a significant increase was estimated in serum creatinine, urea, K⁺, and, urinary Na⁺, total protein, albumin, and urine volume, in obese obestatin treated group. On the other hand, a significant decrease was found in serum Na⁺, total protein, albumin, and, urinary creatinine and K⁺ in obese obestatin treated group in comparison to obese exercised group. These results confirmed that moderate exercise improved kidney function through increasing serum obestatin and decreasing proinflammatory cytokines and improving body metabolism. This was supported by Lin, Lin^[66] and EL-Gohary and Hussien [36] who declared that serum creatinine, urea and albuminuria were declined in obese exercised rats. Also, Park, Kim^[2] confirmed that moderate exercise protected against renal injury in HFD-induced obesity in rats.

Moreover, Toyama, Sugiyama ^[67] stated that regular exercise improved renal function in obesityassociated renal injury in rats and they referred this to elevation of HDL and reduction of advanced glycosylation end products formation. Furthermore, Ishikawa, Gohda ^[68] reported that exercise training decreased albuminuria in obese rats and they owed this to reduction of proinflammatory cytokines. In addition, Gleeson, Bishop ^[69] confirmed the anti-inflammatory action of exercise training and they owed it to visceral fat mass reduction. Therefore, exercise training acted as a natural anti-inflammatory strategy that prevented obesity-induced kidney dysfunction.

On the other hand, Park, Kim^[2] and EL-Gohary and Hussien [36] declared that exercise training improved the antioxidant state and hence improved function. kidnev Also. Aragno. Mastrocola ^[70] found that obestatin had antioxidant activity through increasing activity of prosurvival kinases. Thus, the antioxidant activity of obestatin can be considered as another mechanism by which obestatin improved kidney function changes with obesity. This was confirmed by Koc, Kumral^[71] who reported that obestatin exerted antiinflammatory and antioxidant activities in chronic kidney diseases.

5. Conclusion

This study found that moderate exercise training improved both metabolic and kidney function changes that occurred with HFD-induced obesity through its anti-inflammatory effect and increasing obestatin secretion.

Conflict of Interest

None declared.

References

- Han, T. S., & Lean, M. E. A clinical perspective of obesity, metabolic syndrome and cardiovascular disease, *JRSM Cardiovasc Dis*, 2016; 5, 1-13. doi: 10.1177/2048004016633371.
- 2 Park, S., Kim, C.-S., Lee, J., Kim, J. S., & Kim, J. Effect of regular exercise on the histochemical changes of d-galactose-induced oxidative renal injury in high-fat diet-fed rats,, *Acta Histochem Cytochem*, 2013; 46(4), 111-119. doi: 10.1267/ahc.13012.

- 3 Soltani, Z., Washco, V., Morse, S., & Reisin, E. The impacts of obesity on the cardiovascular and renal systems: cascade of events and therapeutic approaches, *Curr Hypertens Rep*, 2015; 17(2), 7-19. PMID: 25620635.
- Ku, J. M., Andrews, Z. B., Barsby, T., Reichenbach, A., Lemus, M. B. & Drummond, G. R., et al Ghrelin-related peptides exert protective effects in the cerebral circulation of male mice through a nonclassical ghrelin receptor (s), *Endocrinology*, 2015; 156(1), 280-290. doi: 10.1210/en.2014-1415.
- 5 Fernández-Sánchez, A., Madrigal-Santillán, E., Bautista, M., Esquivel-Soto, J., Morales-González, Á. & Esquivel-Chirino, C., et al Inflammation, oxidative stress, and obesity, *Int J Mol Sci*, 2011; 12(5), 3117-3132. doi: 10.3390/ijms12053117.
- 6 Teixeira-Lemos, E., Nunes, S., Teixeira, F., & Reis, F. Regular physical exercise training assists in preventing type 2 diabetes development: focus on its antioxidant and antiinflammatory properties, *Cardiovascular Diabetology*, 2011; 10(1), 12-23. doi: 10.1186/1475-2840-10-12.
- 7 Granata, R., Gallo, D., Luque, R. M., Baragli, A., Scarlatti, F. & Grande, C., et al Obestatin regulates adipocyte function and protects against diet-induced insulin resistance and inflammation, *The FASEB Journal*, 2012; 26(8), 3393-3411. doi: 10.1096/fj.11-201343.
- Zhao, C.-M., Furnes, M. W., Stenström, B., Kulseng, B., & Chen, D. Characterization of obestatin-and ghrelin-producing cells in the gastrointestinal tract and pancreas of rats: an immunohistochemical and electronmicroscopic study, *Cell and tissue research*, 2008; 331(3), 575-587. DOI: <u>10.1007/s00441-</u> <u>007-0514-3</u>.
- 9 Shi, J.-B., Guo, Z.-F., Zheng, X., Wang, Z.-B., & Ma, Y.-J. Circulating obestatin is increased in patients with cardiorenal syndrome and positively correlated with vasopressin, *Peptides*, 2012; 38(2), 377-380. doi: 10.1016/j.peptides.2012.08.022.
- 10 Grönberg, M., Tsolakis, A. V., Magnusson, L., Janson, E. T., & Saras, J. Distribution of obestatin and ghrelin in human tissues: immunoreactive cells in the gastrointestinal

tract, pancreas, and mammary glands. *Journal of Histochemistry & Cytochemistry*, 2008; 56(9), 793-801. doi: 10.1369/jhc.2008.951145.

- Green, B. D., & Grieve, D. J. Biochemical properties and biological actions of obestatin and its relevence in type 2 diabetes, *Peptides*, 2018; 100, 249-259. doi: 10.1016/j.peptides.2017.12.006.
- 12 Agnew, A., Calderwood, D., Chevallier, O., Greer, B., Grieve, D., & Green, B. Chronic treatment with a stable obestatin analog significantly alters plasma triglyceride levels but fails to influence food intake; fluid intake; body weight; or body composition in rats, *Peptides*, 2011; 32(4), 755-762. doi: 10.1016/j.peptides.2010.12.005
- 13 Unniappan, S., Speck, M., & Kieffer, T. J. Metabolic effects of chronic obestatin infusion in rats, *Peptides*, 2008; 29(8), 1354-1361. doi: <u>10.1016/j.peptides.2008.03.023</u>.
- 14 Ren, A.-J., Guo, Z.-F., Wang, Y.-K., Wang, L.-G., Wang, W.-Z. & Lin, L., et al Inhibitory effect of obestatin on glucose-induced insulin secretion in rats, *Biochemical and biophysical research communications*, 2008; 369(3), 969-972. doi: 10.1016/j.bbrc.2008.02.146.
- 15 Gao, X.-Y., Kuang, H.-Y., Liu, X.-M., Wang, X.-Y., Pan, Y.-H., & Ma, X.-X. Decreased obestatin in plasma in metabolically obese, normal-weight men with normal glucose tolerance, *Diabetes research and clinical practice*, 2008; 79(1), e5-e6. doi: 10.1016/j.diabres.2007.07.008.
- 16 Lippl, F., Erdmann, J., Lichter, N., Tholl, S., Wagenpfeil, S., Adam, O., & Schusdziarra, V. Relation of plasma obestatin levels to BMI, gender, age and insulin, *Hormone and metabolic research*, 2008; 40(11), 806-812. doi: 10.1055/s-2008-1081503.
- 17 Egido, E. M., Hernández, R., Marco, J., & Silvestre, R. A. Effect of obestatin on insulin, glucagon and somatostatin secretion in the perfused rat pancreas, *Regulatory peptides*, 2009; 152(1-3), 61-66. doi: <u>10.1016/j.regpep.2008.08.003</u>.
- 18 Zhang, M., Li, F., & Wang, J. Correlation analysis of serum obestatin expression with insulin resistance in childhood obesity, *Genet*

Mol Res, 2017; 28(16), 2-17. doi: <u>10.4238/gmr16029210</u>.

- 19 Granata, R., Settanni, F., Gallo, D., Trovato, L., Biancone, L. & Cantaluppi, V., et al Obestatin promotes survival of pancreatic β -cells and human islets and induces expression of genes involved in the regulation of β -cell mass and function, *Diabetes*, 2008; doi: 10.2337/db07-1104.
- 20 Pradhan, G., Wu, C.-S., Lee, J. H., Kanikarla, P., Guo, S. & Yechoor, V. K., et al Obestatin stimulates glucose-induced insulin secretion through ghrelin receptor GHS-R, *Scientific reports*, 2017; 7(1), 979-993. doi: 10.1038/s41598-017-00888-0.
- 21 Nagaraj, S., Raghavan, A. V., Rao, S. N., Manjappara, U. V. Obestatin and Nt8U influence glycerolipid metabolism and PPAR gamma signaling in mice, *j.biocel*, 2014, 53, 414-422. doi: 10.1016/j.biocel.2014.06.006.
- Gao, X.-Y., Kuang, H.-Y., Liu, X.-M., & Ma, Z.-B. Decreased gastric body mucosa obestatin expression in overweight and obese patients, *Peptides*, 2010; 31(2), 291-296. doi:10.1016/j.peptides.2009.11.001.
- 23 Shen, C., Yu, T., Tang, Z. H., & Wu, K. M. Changes in ghrelin and obestatin levels before and after a meal in children with simple obesity and anorexia, *Hormone research in paediatrics*, 2013; 79(6), 341-346. doi: 10.1159/000351464.
- Wang, W.-M., Li, S.-M., Du, F.-M., Zhu, Z.-C., Zhang, J.-C., & Li, Y.-X. Ghrelin and obestatin levels in hypertensive obese patients, *Journal of International Medical Research*, 2014; 42(6), 1202-1208. doi: 10.1177/0300060514543040.
- 25 Lee, W.-J., Chen, C.-Y., Ser, K.-H., Chong, K., Chen, S.-C. & Lee, P.-C., et al Differential influences of gastric bypass and sleeve gastrectomy on plasma nesfatin-1 and obestatin levels in patients with type 2 diabetes mellitus, *Current Pharmaceutical Design*, 2013; 19(32), 5830-5835. PMID: 23768444.
- 26 Arrigo, T., Gitto, E., Ferrau, V., Munafò, C., Alibrandi, A., Marseglia, G. L. Effect of weight reduction on leptin, total ghrelin and obestatin concentrations in prepubertal children, *Journal of Biological Regulators*

and Homeostatic Agent, 2012; 26(1), S95-S103. PMID:22691243.

- 27 Prodam, F., Cadario, F., Bellone, S., Trovato, L. & Moia, S., et al Obestatin levels are associated with C-peptide and antiinsulin antibodies at the onset, whereas unacylated and acylated ghrelin levels are not predictive of long-term metabolic control in children with type 1 diabetes, *The Journal of Clinical Endocrinology & Metabolism*, 2014; 99(4), E599-E607. doi: 10.1210/jc.2013-3294.
- Siejka, A., Jankiewicz-Wika, J., Kołomecki, K., 28 Cywiński, Piestrzeniewicz, J., K.. Swiętosławski, J. & Komorowski, J. J. C. Long-term impact of vertical banded gastroplasty (VBG) on plasma concentration of soluble leptin receptor, leptin. ghrelin. omentin-1, obestatin, and retinol binding protein 4 (RBP4) in patients with severe obesity, Cvtokine, 2013; 64(2), 490-493. doi: 10.1016/j.cyto.2013.07.026.
- Gibb, A. A., Epstein, P. N., Uchida, S., Zheng, Y., McNally, L. A., Obal, D. & Hill, B. G. Exercise-Induced Changes in Glucose Metabolism Promote Physiological Cardiac Growth, *Circulation*, 2017; 136(22), 2144-2157. doi: 10.1161/CIRCULATIONAHA.117.028274.

30 Mony, A., Batmanabane, M., Hussein, H. H., Mahmoud, O. M., Ukwenya, V., & Ashaolu, O. Effect of obestatin on body weight, serum glucose and insulin levels in albino rats, *Eur. j. anat*, 2013; 17(2), 59-62.

- Alaleem, A., Dalia, I., Elmotteleb, A., & Dalia, M. Possible Involvement of Visfatin in the Beneficial Effect of Simvastatin plus Metformin in High Fat Diet Fed Rats, *Al-Azhar Medical Journal*, 2013; 42(1), 39-56. doi: 10.12816/0015743.
- Novelli, E., Diniz, Y., Galhardi, C., Ebaid, G., Rodrigues, H. & Mani, F., et al Anthropometrical parameters and markers of obesity in rats. *Laboratory animals*, 2007; 41(1), 111-119. doi: 10.1258/002367707779399518.
- 33 D. A. Silva ND, J., Fernandes, T., Soci, U. P., Monteiro, A. W. & Phillips, M. I. Swimming training in rats increases cardiac MicroRNA-126 expression and angiogenesis, *Med Sci*

Sports Exerc, 2012; 44(8), 1453-62. doi: 10.1249/MSS.0b013e31824e8a36.

- 34 Feng, R., Wang, L., Li, Z., Yang, R., Liang, Y. & Sun, Y., et al A systematic comparison of exercise training protocols on animal models of cardiovascular capacity, *Life Sciences*, 2018; 217, 128-140. doi: <u>10.1016/j.lfs.2018.12.001</u>.
- 35 Röhling, M., Herder, C., Stemper, T., & Müssig, K. Influence of Acute and Chronic Exercise on Glucose Uptake, *Journal of diabetes research*, 2016; 2016, 1-33. doi: 10.1155/2016/2868652.
- 36 EL-Gohary, O. A., & Hussien, N. I. J. Effect of exercise and quercetin on obesity induced metabolic and renal impairments in albino rats, *J Phys Pharm Adv*, 2015; 5, 589-602. doi: 10.3109/03009742.2015.1021376.
- 37 Sun, G., Bishop, J., Khalili, S., Vasdev, S., Gill, V., Pace, D. & Zhang, H. J. Serum visfatin concentrations are positively correlated with serum triacylglycerols and down-regulated by overfeeding in healthy young men, *The American Journal of Clinical Nutrition*, 2007; 85(2), 399-404. doi: <u>10.1093/ajcn/85.2.399</u>.
- Wallace, T. M., Levy, J. C., & Matthews, D. R. Use and Abuse of HOMA Modeling, *Diabetes care*, 2004; 27(6), 1487-1495. PMID: 15161807.
- 39 Friedewald, W. T., Levy, R. I., & Fredrickson, D. S. Estimation of the concentration of lowdensity lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge, *Clinical chemistry*, 1972; 18(6), 499-502. PMID: 4337382.
- 40 Kayamori, F., & Igarashi, K. Effects of dietary nasunin on the serum cholesterol level in rats, *Bioscience, biotechnology, and biochemistry*, 1994; 58(3), 570-571. doi: 10.1271/bbb.58.570.
- 41 Noeman, S. A., Hamooda, H. E. & Baalash, A. A. Biochemical study of oxidative stress markers in the liver, kidney and heart of high fat diet induced obesity in rats, *Journal Of Diabetes*, 2011; 3(1), 17-35. doi: 10.1186/1758-5996-3-17.
- 42 Murray, R. Creatinine. Clinical Chemistry theory, analysis, and correlation. Kaplan LA, Pesce AJ, eds. second ed. St. Louis: *The CV Mosby Company*, 1984; p. 1247-1253. doi: 10.12691/ajfn-3-6-1.

- 43 Chow, L. S., Greenlund, L. J., Asmann, Y. W., Short, K. R., McCrady, S. K., Levine, J. A., & Nair, K. S. Impact of endurance training on murine spontaneous activity, muscle mitochondrial DNA abundance, gene transcripts, and function, *Journal of Applied Physiology*, 2007; 102(3), 1078-1089. Doi: 10.1152/japplphysiol.00791.2006.
- Lacquaniti, A., Donato, V., Chirico, V., Buemi, A., Buemi, M. Obestatin: an interesting but controversial gut hormone, *Ann Nutr Metab*, 2011; 59(2-4), 193-199. doi: 10.1159/000334106.
- 45 Wang, Y., Chen, X., Song, Y., Caballero, B., & Cheskin, L. Association between obesity and kidney disease: a systematic review and meta-analysis, *Kidney international*, 2008; 73(1), 19-33. doi: 10.1038/sj.ki.5002586.
- 46 El Sawy, S. A., El-Sherbiny, R. A., El-Saka, M. H., & El-Shaer, R. A. Effect of obestatin on normal, diabetic, and obese male albino rats, *Tanta Medical Journal*, 2016; 44(1), 16-35. doi: 10.4103/1110-1415.180550.
- 47 Saengsirisuwan, V., Perez, F. R., Kinnick, T. R., & Henriksen, E. J. Effects of exercise training and antioxidant R-ALA on glucose transport in insulin-sensitive rat skeletal muscle, *Journal of Applied Physiology*, 2002; 92(1), 50-58. doi: 10.1152/japplphysiol.000617.2001.
- 48 Savini, I., Catani, M., Evangelista, D., Gasperi, V., & Avigliano, L. Obesity-associated oxidative stress: strategies finalized to improve redox state, *Int. J. Mol. Sci.*, 2013; 14(5), 10497-10538. doi: 10.3390/ijms140510497.
- 49 EL-Gohary, O. A. Exercise Training Prevents Age-induced Insulin Resistance in Rats: Effect on Circulating Catecholamines, Inflammatory Cytokines and Skeletal Muscle Glut4 Transporters, *Am. J. Biomed. Sci.*, 2017; 9(3):127-138. doi:10.5099/aj170300127.
- 50 Dewal, R. S. & Stanford, K. I. Effects of exercise on brown and beige adipocytes, Biochimica et *Biophysica Acta (BBA) -Molecular and Cell Biology of Lipids*, 2018; 1864 (1), 71-78. doi: 10.1016/j.bbalip.2018.04.013.
- 51 Mafra, D., Guebre-Egziabher, F., & Fouque, D. J. Endocrine role of stomach in appetite

regulation in chronic kidney disease: about ghrelin and obestatin, *Journal of Renal Nutrition*, 2010; 20(2), 68-73. doi: 10.1053/j.jrn.2009.08.002.

- 52 Seim, I., Walpole, C., Amorim, L., Josh, P., Herington, A., & Chopin, L. The expanding roles of the ghrelin-gene derived peptide obestatin in health and disease, *Molecular and cellular endocrinology*, 2011; 340(1), 111-117. doi: 10.1016/j.mce.2011.03.018.
- 53 Favaro, E., Granata, R., Miceli, I., Baragli, A., Settanni, F., Perin, P. C. & Zanone, M. The ghrelin gene products and exendin-4 promote survival of human pancreatic islet endothelial cells in hyperglycaemic conditions, through phosphoinositide 3-kinase/Akt, extracellular signal-related kinase (ERK) 1/2 and cAMP/protein kinase A (PKA) signalling pathways, *Diabetologia*, 2012; 55(4), 1058-1070. doi: 10.1007/s00125-011-2423-y.
- 54 Nagaraj, S., Peddha, M. S. & Manjappara, U. V. Fragments of obestatin as modulators of feed intake, circulating lipids, and stored fat, *Biochemical and Biophysical Research Communications*, 2008; 366(3), 731-737. doi: 10.1016/j.bbrc.2007.12.036.
- 55 Ivanova, E. A., Myasoedova, V. A., Melnichenko, A. A., Grechko, A. V., & Orekhov, A. N. Small Dense Low-Density Lipoprotein as Biomarker for Atherosclerotic Diseases. Oxidative medicine and cellular longevity, *Oxidative Medicine and Cellular Longevity*, 2017; 2017, 1273042-1273050. doi: 10.1155/2017/1273042.
- 56 Speretta, G. F. F., Rosante, M. C., Duarte, F. O., Leite, R. D., Lino, A. D. d. S., Andre, R. A. & Duarte, A. C. The effects of exercise modalities on adiposity in obese rats, *Clinics*, 2012; 67(12), 1469-1477. doi: 10.6061/clinics/2012(12)19.
- 57 Amin, K. A., Kamel, H. H. & Eltawab, M. A. Protective effect of Garcinia against renal oxidative stress and biomarkers induced by high fat and sucrose diet, *Lipids in Health and Disease*, 2011; 10(1), 6-19. doi: 10.1186/1476-511X-10-6.
- 58 Wang, J., Chen, C., & Wang, R.-Y. Influence of short-and long-term treadmill exercises on levels of ghrelin, obestatin and NPY in plasma

and brain extraction of obese rats, *Endocrine*, 2008; 33(1), 77-83. doi: <u>10.1007/s12020-008-9056-z</u>.

- 59 Kume, S., Uzu, T., Araki, S.-i., Sugimoto, T., Isshiki, K., Chin-Kanasaki, M. & Kadowaki, T. J. Role of altered renal lipid metabolism in the development of renal injury induced by a highfat diet, *Journal of the American Society of Nephrology*, 2007; 18(10), 2715-2723. doi: 10.1681/ASN.2007010089.
- 60 Choudhary, M. I., Naheed, S., Jalil, S., & Alam, J. J. Effects of ethanolic extract of Iris germanica on lipid profile of rats fed on a high-fat diet, *Journal of Ethnopharmacology*, 2005; 98(1-2), 217-220. PMID: 15849875.
- 61 Tokuyama, H., Wakino, S., & Ito, H. Obesity in CKD, Nefrologia, 2008; 66(9), 1770-1777.
- 62 Serra, A., Romero, R., Lopez, D., Navarro, M., Esteve, A., Perez, N. & Ariza, A. Renal injury in the extremely obese patients with normal renal function, *Kidney International*, 2008; 73(8), 947-955. doi: 10.1038/sj.ki.5002796.
- 63 El-Wakf, A. M., Serag, H. M., & Omar, A. J., Alleviating effect of Bauhinia variegata leaves extract on altered serum adipokines and impaired kidney function in male rats with experimentally induced obesity, *J Am Sci*, 2014; 10, 5-14.
- 64 Darouich, S., Goucha, R., Jaafoura, M. H., Zekri, S., Maiz, H. B., & Kheder, A. Clinicopathological characteristics of obesityassociated focal segmental glomerulosclerosis, *Ultrastructural pathology*, 2011; 35(4), 176-182. doi: 10.3109/01913123.2011.584657.
- 65 Habibi, J., Hayden, M. R., Sowers, J. R., Pulakat, L., Tilmon, R. D., Manrique, C. & Whaley-Connell, A. Nebivolol attenuates redox-sensitive glomerular and tubular mediated proteinuria in obese rats, *Endocrinology*, 2010; 152(2), 659-668. doi: 10.1210/en.2010-1038.
- 66 Lin, Q. Q., Lin, R., Ji, Q. L., Zhang, J. Y., Wang, W. R., Yang, L. N., & Zhang, K. F. Effect of exercise training on renal function and renal aquaporin - 2 expression in rats with chronic heart failure, *Clinical and Experimental Pharmacology and Physiology*, 2011; 38(3), 179-185. doi: 10.1111/j.1440-1681.2011.05481.x..

- 67 Toyama, K., Sugiyama, S., Oka, H., Sumida, H., & Ogawa, H. Exercise therapy correlates with improving renal function through modifying lipid metabolism in patients with cardiovascular disease and chronic kidney disease, *Journal of cardiology*, 2010; 56(2), 142-146. doi: 10.1016/j.jjcc.2010.06.007.
- 68 Ishikawa, Y., Gohda, T., Tanimoto, M., Omote, K., Furukawa, M., Yamaguchi, S. & Funabiki, K. Effect of Exercise on Kidney Function, Oxidative Stress, and Inflammation in Type 2 Diabetic KK-A y Mice, *Experimental Diabetes Research*, 2012; 2012, 1-10. doi: 10.1155/2012/702948.
- 69 Gleeson, M., Bishop, N. C., Stensel, D. J., Lindley, M. R., Mastana, S. S., & Nimmo, M. A. The anti-inflammatory effects of exercise: mechanisms and implications for the

prevention and treatment of disease, *Nature Reviews Immunology*, 2011; 11(9), 607-619. doi: 10.1038/nri3041.

- Aragno, M., Mastrocola, R., Ghé, C., Arnoletti, E., Bassino, E., Alloatti, G., & Muccioli, G. Obestatin induced recovery of myocardial dysfunction in type 1 diabetic rats: underlying mechanisms, *Cardiovascular Diabetology*, 2012; 11(1), 129-143. doi: <u>10.1186/1475-2840-11-129</u>.
- 71 Koç, M., Kumral, Z. N. Ö., Özkan, N., Memi, G., Kaçar, Ö. & Bilsel, et al Obestatin improves ischemia/reperfusion-induced renal injury in rats via its antioxidant and antiapoptotic effects: Role of the nitric oxide, *Peptides*, 2014; 60, 23-31. doi: 10.1016/j.peptides.2014.07.019.